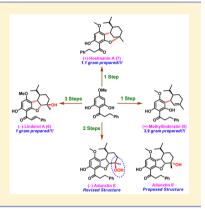
Enantioselective Total Syntheses of (+)-Hostmanin A, (–)-Linderol A, (+)-Methyllinderatin and Structural Reassignment of Adunctin E

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Supporting Information

ABSTRACT: A one-step protocol has been developed for the enantioselective synthesis of hexahydrodibenzofuran derivatives using a modified Friedel–Crafts reaction. The developed method was applied to the synthesis of a series of natural products including (+)-hostmanin A, (+)-methyllinderatin, and (-)-linderol A. The synthetic and spectroscopic data investigations led to the structural reassignment of natural product adunctin E, which was further confirmed by single-crystal X-ray analysis.



INTRODUCTION

The leaves of *Piper adunctum* (Piperaceae), distributed throughout tropical America, have been employed as a folk medicine for the treatment of wounds, dysentery, and diarrhea and as a hemostatic agent. Investigation of the extract of leaves of *P. adunctum* by Orjala et al.¹ led to the isolation of five natural products adunctin A-E.¹ The structures of adunctin $A-E^1$ were confirmed by 1-D and 2-D NMR spectroscopy. The structure of adunctin B (1) was further established by single-crystal X-ray analysis. As shown in Figure 1, adunctin C (2) and

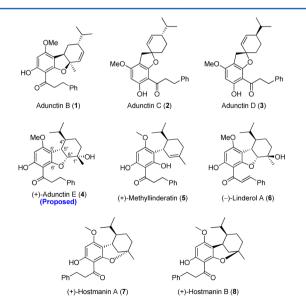


Figure 1. Adunctins and related natural products.

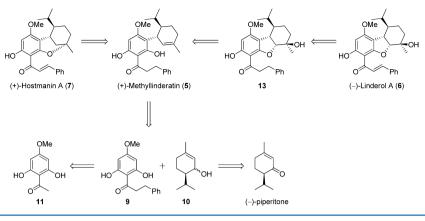
D (3) were found to contain a spirocyclic ring system, while adunctin B (1) and E (4) were novel cinnamoyl-hexaydrodibenzofuran derivatives. Another set of natural products, namely, methyllinderatin (5) and linderol A (6), which have structures similar to those of adunctins, were isolated from the fresh bark of *Lindera umbellata* by K. Ichino² and Mimaki et al.,³ respectively. In 2007, Portet et al.⁴ isolated hostmanin A (7) and B (8) along with known methylinderatin (5), adunctin E (4), and related natural products from the leaves of *Piper hostmannianum*. The structures of hostmanin A (7) and B (8) were confirmed by single-crystal X-ray analysis. Adunctin B (1), C (2), D (3), and methyllinderatin (5) showed antibacterial effects toward *M. luteus* at concentrations of 3.5, 2.4, and 2.5 μ g, respectively.

(–)-Methyllinderatin (**5**) also showed potent antiplasmodial activity with an IC₅₀ value of 5.64 μ M against chloroquine sensitive and resistant strains of *Plasmodium falciparum* (F32, FcB1). The activity of (–)-methyllinderatin (**5**) was confirmed in vivo against *Plasmodium vinckeipetteri* in mice (80% of reduction of parasitemia) at a dose of 20 mg/kg/day.⁴ (–)-Linderol A (**6**) exhibited potent inhibitory activity on melanin biosynthesis of cultured B-16 melanoma cells without causing any cytotoxicity in the cultured cells or skin irritation in guinea pigs.³ Adunctin B (**1**), E (**4**), and linderol A (**6**) have four contiguous stereocenters at 1″, 4″, 5″, and 6″ positions. Interesting structural features coupled with potent biological activities of adunctins and linderols have proven to be a fertile ground for total synthesis.⁵

In 2001, Ohta and co-workers^{5a} reported the first total synthesis of (\pm) linderol A (6) using a coumarin rearrangement

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Scheme 1. Retrosynthetic Analysis



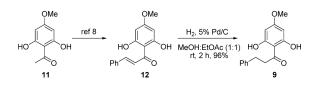
strategy in a 20 step long linear sequence, followed by an asymmetric version by a chiral auxiliary approach.^{5f} Later, in 2011, Yamashita and co-workers^{5j,k} reported the first total synthesis of (\pm) -adunctin B (1) in a 15 step long linear sequence. In 2007, Ohta and co-workers^{5g} reported the total synthesis of the proposed structure of adunctin E (4); however, the spectral data of this compound, including ¹H and ¹³C NMR spectra, were inconsistent with the literature data for adunctin E (4),¹ thus proving that the proposed structure of adunctin E (4) was incorrect.

Most of the previously reported syntheses⁵ of adunctin B (1) and linderol A (6) are racemic, tedious (15-20 step long linear sequences) and are not stereoselective. Herein, we report protecting group free,⁶ concise, and gram-scale enantioselective total syntheses of (+)-methyllinderatin (5), (+)-hostmanin A (7), and (-)-linderol A (6) and the structural reassignment of adunctin E (4).

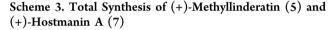
RESULT AND DISCUSSION

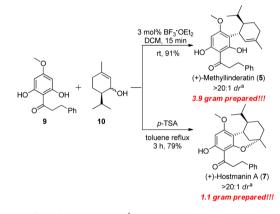
In our quest to synthesize these natural products, we decided to employ the highly diastereoselective modified Friedel-Crafts reaction' that involved two required components, dihydrochalcone derivative 9 and alcohol 10, as shown retrosynthetically in Scheme 1. This route had the advantage of optimum convergency, and it was presumed that the stereochemical outcome of the Friedel-Crafts coupling reaction would be governed by the bulky isopropyl group at the C-4" position. The required chalcone derivative 9 was synthesized from commercially available acetophenone derivative 11 in 4 steps. Thus, 11 was converted to α,β unsaturated ketone derivative 12 in 3 steps by a known protocol,⁸ followed by reduction of the double bond of 12 by 5% Pd/C, under a hydrogen atmosphere to afford the required dihydrochalcone 9 in 96% yield (Scheme 2). The piperetol (10) was obtained by reduction of the keto group of (-)-piperitone using Luche reduction.⁹ With required fragments in hand, the stage was set to investigate the key Friedel-Crafts coupling reaction. Treatment of a mixture of 9 and 10 in the presence of 3 mol % BF3. OEt2 led to rapid

Scheme 2. Preparation of Starting Material



formation of highly regio- and diastereoselective coupling product methyllinderatin (5) in 91% yield with a >20:1 diastereomeric ratio (only major diastereomer is shown in Scheme 3). The spectral data (1 H, 13 C, IR, and HRMS) and



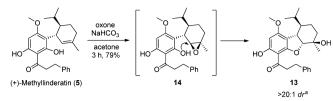


 $^{a}dr = anti/syn$ determined via ¹H NMR spectroscopy.

optical rotation {[α]_D +41.2 (*c* 0.4, CHCl₃)} were in complete agreement with those of natural (+)-methyllinderatin (5)² {lit² [α]_D +41.0 (*c* 0.4, CHCl₃)}. The one step synthesis of (+)-Hostmanin A (7) was also achieved. Thus, the reaction of dihydrochalcone derivative 9 with alcohol 10 in the presence of 30 mol % *p*-TSA in refluxing toluene furnished (+)-hostmanin A (7) as a single diastereomer with 79% yield, whose spectral data (¹H, ¹³C, IR, and HRMS) and optical rotation {[α]_D +59.4 (*c* 0.15, MeOH)} were identical to those reported for the natural product⁴ (Scheme 3).

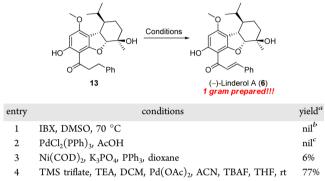
Next, we turned our attention toward the total synthesis of (-)-linderol A (6). Treatment of DMDO with (+)-methyllindratin (5) directly generated the advanced tricyclic intermediate 13 in 79% yield and >20:1 diastereomeric ratio (determined by crude ¹H NMR) by epoxidation of the double bond, followed by opening of epoxide 14 by the adjacent phenolic-OH group, as shown in Scheme 4. Conversion of tricyclic intermediate 13 to linderol A (6) required the formation of a double bond at the benzylic position. After trying various conditions, as shown in Table 1, we were finally successful in achieving the enantioselective total synthesis of linderol A (6). Thus, compound 13 under Saegusa–Ito oxidation condition¹⁰ gave (-)-linderol A (6) in 77% yield. The spectral data (¹H, ¹³C, IR,

Scheme 4. Toward the Total Synthesis of (-)-Linderol A (6)



 $^{a}dr = syn/anti$ determined via ¹H NMR spectroscopy.

Table 1. Screening of Conditions for $\alpha_{,\beta}$ Unsaturation of 13



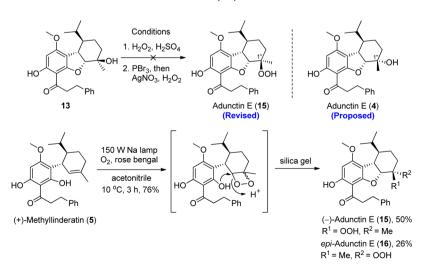
^{*a*}Isolated yields. ^{*b*}Decomposition of starting material. ^{*c*}Recovered starting material.

and HRMS) and optical rotation { $[\alpha]_D - 23.4$ (*c* 0.42, CHCl₃)} of synthetic linderol A (6) were in complete agreement with those of natural linderol A (6).³ {lit³ $[\alpha]_D - 22.7$ (CHCl₃)}.

After completing the enantioselective total syntheses of (+)-methyllindratin (5), (+)-hostmanin A (7), and (-)-linderol A (6), we next turned out attention toward the structural revision of adunctin E (4) and the total synthesis of the revised structure. As discussed earlier, Ohta and co-workers^{5g} reported that the proposed structure of adunctin E (4) by Sticher group¹ was incorrect. After careful comparison of ¹H and ¹³C NMR spectral data, we found that the major difference is that adunctin E (4) has a doublet at δ 4.5 in ¹H and tertiary C-1" at 80.9 ppm. The values for dihydrolinderol **13** (prepared by our group) and the proposed structure of adunctin E (4) (prepared by Ohta and co-workers)^{5g} are at δ 4.13 and 4.26 in ¹H and 69.5 and 70.4 ppm, respectively, in ¹³C NMR. This made us to think that, most probably, structural revision needs to be done at C-1" and or C-6", as the deshielding of the C-6" proton by ~0.3 ppm in ¹H and C-1" by ~11 ppm in ¹³C was observed. After going through various literature reports and ¹H and ¹³C NMR data of related compounds (e.g., artemisinin, plakortolides, etc.),¹¹ it was observed that a tertiary carbon containing peroxide linkage also comes in the range of 78-84 in ¹³C NMR. On the basis of the above observation, we proposed the revised structure of the adunctin E as hydroperoxide 15, as shown in Scheme 5. To confirm this, we attempted the synthesis of the revised structure of adunctin E(15). Our initial attempt to convert the hydroxyl group of compound 13 to hydroperoxide by using H₂O₂ under acidic conditions as well as by using PBr₃, followed by H_2O_2 in the presence of AgNO₃, was unsuccessful. Next, we relied on [2 + 2] cycloaddition of (+)-methyllinderatin (5) with singlet oxygen, followed by a ring-opening protocol. Thus, [2 + 2] cycloaddition reaction of (+)-methyllinderatin (5) with singlet oxygen using a sodium lamp in the presence of rose bengal directly afforded a diastereomeric mixture of hydroperoxides 15 and 16 in a 2:1 ratio with 76% yield, which was carefully separated by silica gel column chromatography. The spectral data (¹H, ¹³C, IR, and HRMS) of major isomer 15 was in complete agreement with the reported data,¹ but the sign of the optical rotation $\{[\alpha]_{D}\}$ -17.1 (c 0.65, MeOH)} {lit¹ [α]_D +16.3 (c 0.65, MeOH)} was exactly opposite, confirming that synthetic 15 is an antipode of the natural (+)-adunctin E.¹ The stereochemistry of (-)-adunctin E (15) was unambiguously established by single-crystal X-ray analysis (Figure 2),¹² which established the absolute configuration of natural (+)-adunctin E (15) as (4''S, 5''R, 6''S, 1''S). It is worth mentioning that the stereochemistry of tertiary carbon, containing peroxide (C-1'') was found to be exactly opposite to that of the proposed structure of adunctin E (4) (see Scheme 5). Using the same strategy, natural (+)-adunctin E (15) could be synthesized from dihydrochalcone derivative 9 and ent-10. Thus, the total synthesis of the proposed structure of adunctin E (4) by Ohta and co-workers, ^{5g} followed by synthesis of 13, 15, and 16 by our group, led to an unambiguous reassignment of the structural composition and established the relative and absolute stereochemical configuration of the natural product adunctin E.

In conclusion, we achieved concise, protecting group free, gram-scale, highly atom economic, and enantioselective

Scheme 5. Total Synthesis of Revised Structure of Adunctin E (15)



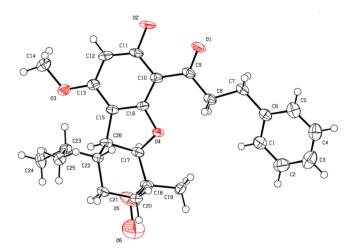


Figure 2. ORTEP diagram of (-)-adunctin E (15). Thermal ellipsoids are drawn at the 50% probability level.

syntheses of four natural products, namely, (+)-methyllinderatin, (+)-hostmanin A, (-)-linderol A, and (-)-adunctin E, using a modified Friedel–Crafts reaction, and a photochemical [2 + 2] cycloaddition of olefin with singlet oxygen as key steps. The synthetic approach allows ready access to analogues that can be used for further biological studies.

EXPERIMENTAL SECTION

General Information. All reactions were carried out under a nitrogen atmosphere with dry solvents under anhydrous conditions, unless otherwise mentioned. All the chemicals were purchased commercially and used without further purification. Anhydrous THF and diethyl ether were distilled from sodium benzophenone, and dichloromethane was distilled from calcium hydride. Yields refer to chromatographically pure compounds, unless otherwise stated. Reactions were monitored by thin-layer chromatography (TLC) carried out on 0.25 mm silica gel plates (60F-254) using UV light as a visualizing agent and a p-anisaldehyde or ninhydrine stain, and heat as developing agents. Silica gel (particle size: 100-200 and 230-400 mesh) was used for flash column chromatography. Neat coumpounds were used for recording IR spectra. NMR spectra were recorded on either 400 (¹H, 400 MHz; ¹³C, 100 MHz) or 500 (¹H, 500 MHz; ¹³C, 125 MHz). Mass spectrometric data were obtained using Q-Tof-Premier-HAB213 and Q-Tof-Premier-ESI-MS instruments. Melting points measurements were made using a hot stage apparatus. Optical rotations were measured using a polarimeter at 20 °C.

The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublet, ddd = doublet of a doublet of a doublet, dt = doublet of a triplet, td = triplet of a doublet, m = multiplet, br = broad.

Experimental Procedures. 1-(2,6-Dihydroxy-4-methoxyphenyl)-3-phenylpropan-1-one (9). To a magnetically stirred solution of 12 (6 g, 22.2 mmol) in methanol (30 mL) and ethyl acetate (30 mL) was added 5% Pd/C (400 mg) at room temperature. The resulting mixture was stirred at rt for 2 h under H₂ bladder pressure. After completion of reaction, indicated by TLC, the reaction mixture passed through Celite. Evaporation of the solvent and purification of the residue on silica gel column using EtOAc-hexane (1:6) as eluent furnished 9 (5.85 g, 97%) as a pale yellow solid; $R_f = 0.30$ (EtOAchexane 1:4); mp: 173–176 °C; IR (neat): $\nu_{\rm max}/{\rm cm}^{-1}$ 3253 (br.), 2922, 1593, 1526, 1440, 1229, 1081, 810; ¹H NMR (500 MHz, DMSO-*d*₆): δ 12.29 (s, 2H), 7.25-7.20 (m, 4H), 7.16-7.12 (m, 1H), 5.92 (s, 2H), 3.70 (s, 3H), 3.28 (t, J = 8.2 Hz, 2H), 2.87 (t, J = 8.2 Hz, 2H); ¹³C NMR (125 MHz, DMSO-d₆): δ 209.8, 170.8, 169.3, 146.9, 133.6, 131.1, 109.8, 98.5, 60.7, 50.5, 35.3; HRMS: m/z calcd for C₁₆H₁₇O₄ $[M + H]^+$: 273.1127; found: 273.1122. Spectral data were consistent with those previously reported.^{13a}

(*R*)-6-Isopropyl-3-methylcyclohex-2-enol (10). To a magnetically stirred solution of (–)-piperitone (2 g, 13.14 mmol) in MeOH was added CeCl₃·7H₂O (4.89 g, 13.14 mmol), cooled to 0 °C. After 10 min, NaBH₄ (497 mg, 13.14 mmol) was added portionwise for 5 min, and the reaction was allowed to stir at the same temperature. After completion of reaction indicated by TLC, the reaction was quenched by water, and the reaction mixture was concentrated under reduced pressure and then extracted with diethyl ether (2 × 15 mL). Evaporation of the solvent and purification of the residue on silica gel column using EtOAc–hexane (1:49) as eluent furnished 10 (1.78 g, 88%) as a colorless liquid; $R_f = 0.42$ (EtOAc–hexane 1:19). Spectral data were consistent with those previously reported.^{13b}

(+)-Methyllinderatin (5). To a magnetically stirred solution of compound 9 (2.9 g, 10.6 mmol) and 10 (2.14 g, 13.84 mmol) in anhydrous DCM (30 mL) was added BF₃·OEt₂ (40μ L, 0.32 mmol) at room temperature. The resulting reaction mixture was then stirred at rt for 15 min. After completion of reaction indicated by TLC, the reaction was quenched by saturated Na₂CO₃ solution (5 mL) and then extracted with DCM (2 \times 25 mL). Evaporation of the solvent and purification of the residue on silica gel column using EtOAc-hexane (1:49) as eluent furnished methyllinderatin (5) (3.955 g, 91%) as a pale yellow oil; $R_f = 0.60$ (EtOAc-hexane 1:4); $[\alpha]_D^{20}$ +41.2 (c 0.4, CHCl₃); IR (neat): ν_{max}/cm^{-1} 3354 (br.), 2955, 2932, 1624, 1584, 1248, 1212, 816, 698; ¹H NMR (500 MHz, CDCl₃): δ 7.32-7.19 (m, 5H), 7.06 (s, 1H), 6.05 (s, 1H), 5.46 (br. s, 1H), 3.87 (br. d, J = 10.6 Hz, 1H), 3.78 (s, 3H), 3.39 (t, J = 7.6 Hz, 2H), 3.00 (t, J = 7.6 Hz, 2H), 2.17-2.05 (m, 2H), 1.79 (s, 3H), 1.78-1.74 (m, 1H), 1.56-1.50 (m, 1H), 1.47-1.42 (m, 1H), 1.40-1.34 (m, 1H), 0.84 (d, J = 8.0 Hz, 3H), 0.81 (d, J = 8.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 205.3, 165.4, 164.0, 158.9, 142.4, 141.9, 129.0, 128.7, 126.2, 124.7, 109.2, 106.2, 92.3, 55.9, 46.4, 43.8, 35.0, 31.0, 30.9, 28.2, 24.1, 22.4, 22.0, 16.5; HRMS: m/z calcd for $C_{26}H_{33}O_4$ [M + H]⁺: 409.2379; found: 409.2370.

(+)-Hostmanin A (7). To a magnetically stirred solution of compound 9 (1 g, 3.67 mmol) and 10 (736.42 mg, 4.77 mmol) in anhydrous toluene (10 mL) was added p-TSA·H₂O (209 mg, 1.1 mmol) at rt. The resulting reaction mixture was then refluxed at 110 °C for 3 h. After completion of reaction indicated by TLC, the reaction was guenched by a saturated solution of Na₂CO₂ (5 mL) and then extracted with ethyl acetate (2 \times 20 mL). Evaporation of the solvent and purification of the residue on silica gel column using EtOAc-hexane (1:199) as eluent furnished (+)-hostmanin A (7) (1.19 g, 79%) as a pale yellow oil; $R_f = 0.82$ (EtOAc-hexane 1:4); $[\alpha]_{\rm D}^{25}$ +59.4 (c 0.15, MeOH); IR (neat): $\nu_{\rm max}/{\rm cm}^{-1}$ 2928, 1617, 1586, 1216, 1146, 1108; ¹H NMR (400 MHz, CDCl₃): δ 7.32-7.21 (m, 5H), 6.02 (s, 1H), 3.83 (s, 3H), 3.52-3.44 (m, 2H), 3.42-3.36 (m, 1H), 3.06 (t, J = 7.8 Hz, 2H), 1.90 (dd, J = 13.3, 2.1 Hz, 1H), 1.81-1.71 (m, 2H), 1.63 (dd, J = 13.7, 4.6 Hz, 1H), 1.56-1.48 (m, 3H), 1.36 (s, 3H), 1.24–1.19 (m, 1H), 1.07 (d, J = 6.4 Hz, 3H), 0.96 (d, J = 6.4 Hz, 3H); $^{13}\mathrm{C}$ NMR (125 MHz, CDCl₃): δ 204.9, 165.3, 162.2, 159.0, 141.8, 128.4, 128.3, 125.9, 106.8, 104.7, 91.2, 76.8, 55.7, 45.4, 44.0, 35.3, 30.7, 30.1, 29.2, 27.2, 26.3, 22.1, 21.0, 20.5; HRMS: m/z calcd for C₂₆H₃₃O₄ [M + H]⁺: 409.2379; found: 409.2379.

1-((5aR,6R,9R,9aS)-3,6-Dihydroxy-9-isopropyl-1-methoxy-6methyl-5a,6,7,8,9,9a-hexahydrodibenzo[b,d]furan-4-yl)-3-phenylpropan-1-one (13). To a magnetically stirred solution of (+)-methylinderatin (5) (2 g, 4.9 mmol) in acetone (20 mL) was added Na_2CO_3 (1.234 g, 14.68 mmol). The reaction mixture was then cooled to 0 °C and stirred for 10 min. Then, a solution of oxone (6.02 g, 9.8 mmol) (prepared by dissolving oxone in 25 mL water) was added dropwise. The reaction continued at the same temperature for an additional 30 min and then was slowly allowed to come to room temperature. After completion of reaction, indicated by TLC, the reaction mixture was concentrated under reduced pressure and then extracted with ethyl acetate (2 \times 30 mL). Evaporation of the solvent and purification of the residue on silica gel column using EtOAchexane (1:11) as eluent furnished 13 (1.64 g, 79%) as a yellow solid; $R_f = 0.55$ (EtOAc-hexane 1:4); mp: 143–145 °C; $[\alpha]_D^{25}$ –34.9 (c 0.55, ČHCl₃); IR (neat): ν_{max}/cm^{-1} 3484 (br.), 2959, 1633, 1602, 1443, 1374, 1215, 1207, 1083, 977, 812, 718; ¹H NMR (500 MHz, CDCl₃):

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δ 7.29–7.16 (m, 5H), 6.03 (s, 1H), 4.13 (d, *J* = 5.2 Hz, 1H), 3.80 (s, 3H), 3.40–3.30 (m, 2H), 3.09 (dd, *J* = 11.2, 5.4 Hz, 1H), 3.01 (t, *J* = 7.7 Hz, 2H), 2.38 (br. s, 1H), 1.83 (dq, *J* = 6.7, 2.6 Hz, 1H), 1.77–1.72 (m, 1H), 1.66–1.60 (m, 1H), 1.43–1.35 (m, 2H), 1.37 (s, 3H), 1.09–1.03 (m, 1H), 0.88 (d, *J* = 6.9 Hz, 3H), 0.83 (d, *J* = 6.9 Hz, 3H); ¹³C NMR (500 MHz, CDCl₃): δ 203.6, 165.6, 162.1, 141.6, 128.7 (2C), 128.6, 126.3, 113.5, 103.0, 93.1, 92.7, 69.7, 55.8, 46.9, 44.0, 39.9, 35.6, 30.5, 28.4, 27.5, 22.1, 17.5, 15.7; HRMS: *m*/*z* calcd for C₂₆H₃₃O₅ [M + H]⁺: 425.2328; found: 425.2324.

(-)-Linderol A (6). To a magnetically stirred solution of 13 (1.3 g, 3.06 mmol) and TEA (853 µL, 6.12 mmol) in DCM (2 mL) was added TMSOTf (736 µL, 3.98 mmol) dropwise. The resulting solution was stirred at 0 °C for 10 min and then at room temperature for an additional 1 h. The reaction mixture was quenched with a saturated NaHCO3 solution, extracted with DCM, washed with brine, dried over anhydrous Na₂SO₄, and concentrated at reduced pressure. The residue was dissolved in THF (10 mL), and Pd(OAc)₂ (687 mg, 6.12 mmol) was added to the mixture. The reaction mixture was stirred at room temperature for 2 h and then filtered through a Celite pad and washed with EtOAc. The filtrate was concentrated and dissolved in THF (10 mL), and TBAF (3.06 mL, 1 M in THF) was added to this mixture. The mixture was stirred at room temperature for 1 h and then quenched with water, extracted with EtOAc, washed with brine, and dried over anhydrous Na2SO4. Evaporation of the solvent and purification of the residue on silica gel column using EtOAc-hexane (1:11) as eluent furnished the compound (-)-linderol A (6) (1 g, 77%) as a yellow solid; $R_f = 0.50$ (EtOAc-hexane 1:4); mp: 180–182 °C; $[\alpha]_{D}^{20}$ –23.4 (c 0.42, CHCl₃); IR (neat): ν_{max}/cm^{-1} 3446 (br.), 2925, 1639, 1589, 1346, 1031, 809, 458 ; ¹H NMR (500 MHz, CDCl₃): δ 8.09 (d, J = 15.5 Hz, 1H), 7.86 (d, J = 15.5 Hz, 1H), 7.61 (d, J = 4.0 Hz, 2H), 7.42–7.37 (m, 3H), 6.08 (s, 1H), 4.24 (d, J = 5.2 Hz, 1H), 3.84 (s, 3H), 3.14 (dd, J = 11.2, 5.4 Hz, 1H), 1.87-1.81 (m, 2H), 1.77-1.72 (m, 1H), 1.61 (s, 3H), 1.57 (br. s, 1H), 1.45-1.38 (m, 2H), 1.13 (t, J = 11.5 Hz, 1H), 0.91 (d, J = 6.3 Hz, 3H), 0.84 (d, J = 6.9 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 191.1, 166.7, 162.1, 161.5, 143.4, 135.3, 130.3, 128.9, 128.4, 125.8, 113.3, 103.3, 93.0, 92.3, 69.4, 55.5, 46.5, 39.5, 35.4, 28.3, 27.1, 21.8, 17.2, 15.4; HRMS: m/z calcd for C₂₆H₃₁O₅ [M + H]⁺: 423.2171; found: 423.2172.

(-)-Adunctin E (15) and (-)-epi-Adunctin E (16). To a magnetically stirred solution of (+)-methyllinderatin (5) (100 mg, 0.24 mmol) in acetonitrile was added rose bengal (5 mg, 4.9 μ mol). The temperature of reaction was maintained at 10 °C (by putting cold water). The reaction mixture was then continuously purged with an O_2 balloon (by inserting another open needle into the septa of RB) and then exposed to a 150 W sodium lamp. After consumption of starting material indicated by TLC, the sodium lamp was removed and silica gel (230-400 mesh size, 500 mg) was added to the reaction mixture, which was stirred at rt for an additional 1 h. The reaction mixture was then concentrated under reduced pressure and extracted with ethyl acetate $(2 \times 5 \text{ mL})$ and dried over anhydrous Na₂SO₄. Evaporation of the solvent and purification of the residue on silica gel column using monoglyme-hexane (1:7) as eluent furnished (-)-epi-adunctin E (16) (27 mg, 26%) as a white solid; $R_t = 0.55$ (monoglyme-hexane 1:4); mp: 111–112 °C; $[\alpha]_{\rm D}^{20}$ –38 (c 0.2, MeOH); IR (neat): $\nu_{\rm max}$ cm⁻¹ 3384 (br.), 2926, 2874, 1622, 1598, 1358, 1184, 817, 699; ¹H NMR (500 MHz, CDCl₃): δ 7.32-7.16 (m, 5H), 5.99 (s, 1H), 4.45 (d, J = 4.6 Hz, 1H), 3.75 (s, 3H), 3.36-3.29 (m, 2H), 3.00-2.95 (m, 2H), 3.00-2.95 (m, 3.00-2.952H), 2.91 (d, J = 4.0 Hz, 1H), 1.82-1.77 (m, 1H), 1.74-1.66 (m, 1H), 1.63-1.51 (m, 2H), 1.32 (s, 3H), 1.15-1.05 (m, 2H), 0.81 (d, J = 6.3 Hz, 3H), 0.77 (d, J = 6.9 Hz, 3H); ¹³C NMR (500 MHz, CDCl₃): *δ* 203.7, 165.9, 162.0, 161.7, 141.5, 128.9, 128.6, 126.6, 113.2, 103.1, 93.3, 89.5, 82.2, 55.9, 47.2, 44.5, 41.4, 31.0, 30.6, 27.2, 22.1, 21.6, 19.8, 15.6; HRMS: m/z calcd for $C_{26}H_{33}O_6 [M + H]^+$: 441.2277; found: 441.2273. Further elution of the column with monoglymehexane (1:7) gave (-)-adunctin E (15) (54 mg, 50%) as a white solid; $R_{\rm f} = 0.50$ (monoglyme-hexane 1:4); mp: 100-103 °C; $[\alpha]_{\rm D}^{20}$ -17.1 (c 0.65, MeOH); IR (neat): $\nu_{\rm max}/{\rm cm}^{-1}$ 3393 (br.), 2931, 2870, 1631, 1602, 1369, 1205, 813, 698; ¹H NMR (500 MHz, CDCl₃): δ 7.31– 7.20 (m, 5H), 6.04 (s, 1H), 4.49 (d, J = 5.2 Hz, 1H), 3.81 (s, 3H), 3.43–3.30 (m, 2H), 3.08 (dd, J = 11.5, 5.7 Hz, 1H), 3.02 (t, J = 7.7 Hz,

2H), 2.00 (d, J = 14.9 Hz, 1H), 1.86–1.80 (m, 1H), 1.62–1.54 (m, 1H), 1.43 (s, 3H), 1.38–1.30 (m, 2H), 1.08 (t, J = 11.5 Hz, 1H), 0.87 (d, J = 6.9 Hz, 3H), 0.83 (d, J = 6.9 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 203.3, 165.2, 161.8, 161.6, 141.2, 128.4, 128.3, 126.0, 112.9, 102.6, 92.8, 87.6, 80.9, 55.5, 46.3, 43.7, 39.7, 31.9, 30.1, 27.1, 22.0, 21.8, 17.1, 15.4; HRMS: m/z calcd for C₂₆H₃₃O₆ [M + H]⁺: 441.2277; found: 441.2271.

ASSOCIATED CONTENT

S Supporting Information

Copies of ¹H and ¹³C spectra for all new compounds and X-ray crystal data of compound **15** (CIF file). This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) Orjala, J.; Wright, A. D.; Erdelmeier, C. A. J.; Sticher, O.; Raili, T. *Helv. Chim. Acta* **1993**, *76*, 1481–1488.

(2) Ichino, K. Phytochemistry 1989, 28, 955-956.

(3) Mimaki, Y.; Kameyama, A.; Sashida, Y.; Miyata, Y.; Fujii, A. *Chem. Pharm. Bull.* **1995**, 43, 893–895.

(4) Portet, B.; Fabre, N.; Roumy, V.; Gornitzka, H.; Bourdy, G.; Chevalley, S.; Sauvain, M.; Valentin, A.; Moulis, C. *Phytochemistry* **2007**, *68*, 1312–1320.

(5) (a) Yamashita, M.; Ohta, N.; Kawasaki, I.; Ohta, S. Org. Lett. 2001, 3, 1359-1362. (b) Yamashita, M.; Ohta, N.; Shimizu, T.; Matsumoto, K.; Matuura, Y.; Kawasaki, I.; Tanaka, T.; Maezaki, N.; Ohta, S. J. Org. Chem. 2003, 68, 1216-1224. (c) Yamashita, M.; Shimizu, T.; Kawasaki, I.; Ohta, S. Tetrahedron: Asymmetry 2004, 15, 2315-2317. (d) Yamashita, M.; Inaba, T.; Shimizu, T.; Kawasaki, I.; Ohta, S. Synlett 2004, 1897-1900. (e) Yamashita, M.; Inaba, T.; Nagahama, M.; Shimizu, T.; Kosaka, S.; Kawasaki, I.; Ohta, S. Org. Biomol. Chem. 2005, 3, 2296-2304. (f) Yamashita, M.; Shimizu, T.; Inaba, T.; Takada, A.; Takao, I.; Kawasaki, I.; Ohta, S. Heterocycles 2005, 65, 1099-109. (g) Yamashita, M.; Yadav, N. D.; Sawaki, T.; Takao, I.; Kawasaki, I.; Sugimoto, Y.; Miyatake, A.; Murai, K.; Takahara, A.; Kurume, A.; Ohta, S. J. Org. Chem. 2007, 72, 5697-5703. (h) Yamashita, M.; Yadav, N. D.; Sumida, Y.; Kawasaki, I.; Kurume, A.; Ohta, S. Tetrahedron Lett. 2007, 48, 5619-5622. (i) Berber, H.; Delaye, P.-O.; Mirand, C. Synlett 2008, 94-96. (j) Delaye, P.-O.; Lameiras, P.; Kervarec, N.; Mirand, C.; Berber, H. J. Org. Chem. 2010, 75, 2501-2509. (k) Arimitsu, K.; Nomura, S.; Iwasaki, H.; Ozeki, M.; Yamashita, M. Tetrahedron Lett. 2011, 52, 7046-7048. (1) Nomura, S.; Arimitsu, K.; Yamaguchi, S.; Kosuga, Y.; Kakimoto, Y.; Komai, T.; Hasegawa, K.; Nakanishi, A.; Miyoshi, T.; Iwasaki, H.; Ozeki, M.; Kawasaki, I.; Kurume, A.; Ohta, S.; Yamashita, M. Chem. Pharm. Bull. 2012, 60, 94-103.

(6) (a) Young, I. S.; Baran, P. S. Nat. Chem. 2009, 1, 193-205.
(b) Roulland, E. Angew. Chem., Int. Ed. 2011, 50, 1226-1227.
(c) Jansen, D. J.; Shenvi, R. A. Future Med. Chem. 2014, 6, 1127.
(d) Kuttruff, C. A.; Eastgate, M. D.; Baran, P. S. Nat. Prod. Rep. 2014, 31, 419-432.

(7) For early examples in Friedel-Crafts chemistry, see: (a) Price, C. C.; Lund, M. J. Am. Chem. Soc. **1940**, 62, 3105-3107. (b) Burwell, R.

The Journal of Organic Chemistry

L., Jr.; Archer, S. J. Am. Chem. Soc. 1942, 64, 1032–1034.
(c) Streitwieser, A., Jr.; Stang, P. J. J. Am. Chem. Soc. 1965, 87, 4953.
(d) Muhlthau, F.; Stadler, D.; Goeppert, A.; Olah, G. A.; Surya Prakash, G. K.; Bach, T. J. Am. Chem. Soc. 2006, 128, 9668–9675.
(e) Nitsch, D.; Huber, S. M.; Pöthig, A.; Narayanan, A.; Olah, G. A.; Surya Prakash, G. K.; Bach, T. J. Am. Chem. Soc. 2014, 136, 2851–2857.

(8) Chen, Y.-H.; Wang, W.-H.; Wang, Y.-H.; Lin, Z.-Y.; Wen, C.-C.; Chern, C.-Y. *Molecules* **2013**, *18*, 2052–2060.

(9) Gemal, A. L.; Luche, J. L. J. Am. Chem. Soc. 1981, 103, 5454-5459.

(10) Ito, Y.; Hirao, T.; Saegusa, T. J. Org. Chem. 1978, 43, 1011–1013.

(11) (a) Qinghaosu Research Group. Sci. Sin. 1980, 23, 380–396.
(b) Matsumoto, M.; Dobashi, S.; Kuroda, K.; Kondo, K. Tetrahedron 1985, 41, 2147–2154. (c) Capon, R. J.; MacLeod, J. K.; Willis, A. C. J. Org. Chem. 1987, 52, 339–342. (d) O'Shea, K. E.; Foote, C. S. J. Am. Chem. Soc. 1988, 110, 7167–7170. (e) Suzuki, M.; Ohtake, H.; Kameya, Y.; Hamanaka, N.; Noyori, R. J. Org. Chem. 1989, 54, 5292–5302. (f) Rudi, A.; Kashman, Y. J. Nat. Prod. 1993, 56, 1827–1830. (g) Tanaka, J.; Higa, T.; Suwanborirux, K.; Kokpol, U.; Bernardinelli, G.; Jefford, C. W. J. Org. Chem. 1993, 58, 2999–3002. (h) Toth, S. I.; Schmitz, F. J. J. Nat. Prod. 1995, 58, 27–36. (j) Varoglu, M.; Peters, B. M.; Crews, P. J. Nat. Prod. 1995, 58, 27–36. (k) Yong, K. W. L.; Lambert, L. K.; Hayes, P. Y.; De Voss, J. J.; Garson, M. J. J. Nat. Prod. 2012, 75, 351–360.

(12) CCDC 1040618 contains supplementary crystallographic data for (–)-adunctin E (15). These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam. ac.uk/data_request/cif.

(13) (a) Thévenin, M.; Mouray, E.; Grellier, P.; Dubois, J. *Eur. J. Org. Chem.* 2014, 2014, 2986–2992. (b) Wińska, K.; Grudniewska, A.; Chojnacka, A.; Białońska, A.; Wawrzeńczyk, C. *Tetrahedron: Asymmetry* 2010, 21, 670–678.

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